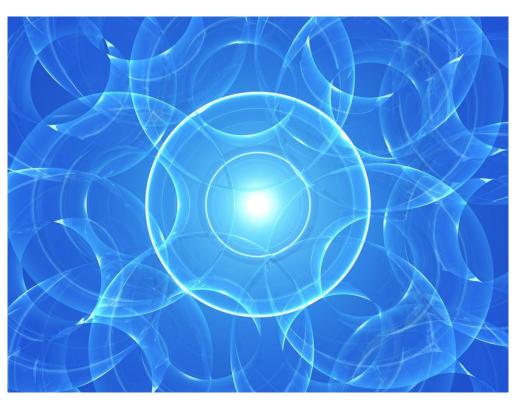


C18, C18-WP, HFC18-16, HFC18-30, RP-AQUA, C8, PFP, Phenyl, HILIC-Amide and 2-EP

HPLC column

SunShell



Core Shell Particle



ChromaNik Technologies Inc.

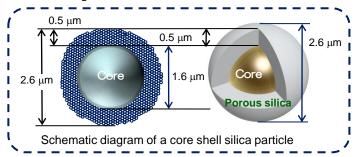


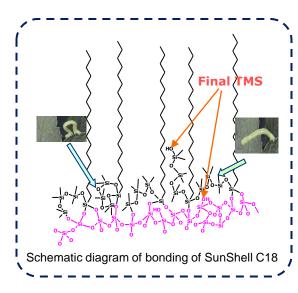
The next generation to Core Shell particle

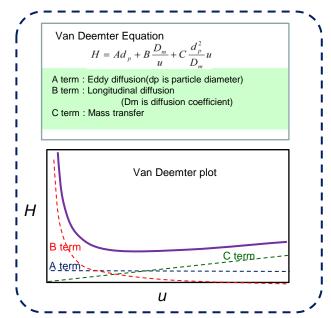
Superficially porous silica

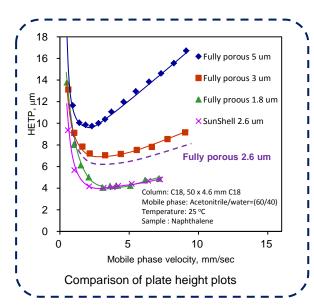
Features of SunShell

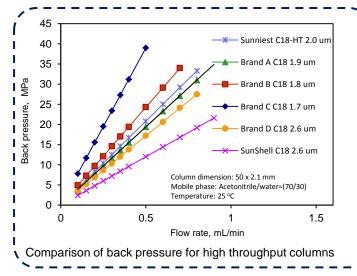
- *1.6 μm of core and 0.5 μm of superficially porous silica layer
- *Same efficiency and high throughput as a Sub 2µm particle
- *Same pressure as a 3 µm particle (less than a half then a sub 2µm particle)
- *Same chemistry as Sunniest technology (reference figure 1)
- *Good peak shape for all compounds such as basic, acidic and chelating compounds
- *High stability (pH range for SunShell C18, 1.5 to 10)
- * Low breeding











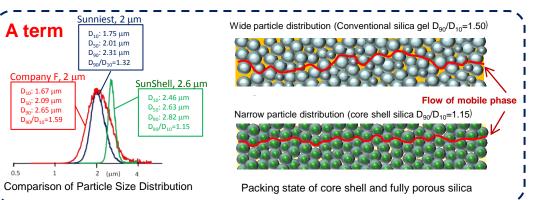
SunShell C18 shows same efficiency as a sub 2 μm C18. In comparison between fully porous 2.6 μm and core shell 2.6 μm (SunShell), SunShell shows lower values for A term, B term and C term of Van Deemter equation. The core shell structure leads higher performance to compare with the fully porous structure.

Furthermore back pressure of SunShell C18 is less than a half to compare with sub-2 μm C18s.



Why does a 2.6 μm core shell particle show the same performance as a sub 2 μm particle?

All terms in Van Deemter Equation reduce.

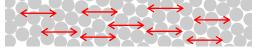


The size distribution of a core shell (SunShell) particle is much narrower than that of a conventional totally porous particle, so that the space among particles in the column reduces and efficiency increases by reducing Eddy Diffusion (multi-path diffusion) as the A term in Van Deemter Equation.

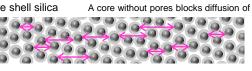
Diffusion of a solute is blocked by the existence of a core, so that a solute diffuses less in a core shell silica column than in a totally porous silica column. Consequently B term in Van Deemter Equation reduces in the core shell silica column.

B term

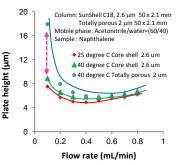
Totally porous silica A solute diffuses in a pore as well as outside of particles.



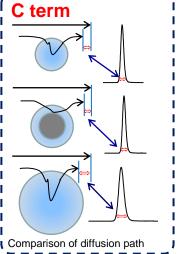
Core shell silica A core without pores blocks diffusion of a solute.



Difference of longitudinal diffusion

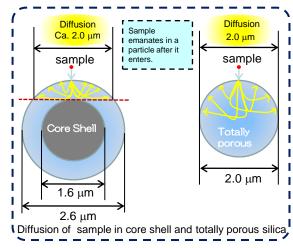


Plot of Flow rate and Plates height



As shown in the left figure, a core shell particle has a core so that the diffusion path of samples shortens and mass transfer becomes fast. This means that the C term in Van Deemter Equation reduces. In other words, HETP (theoretical plate) is kept even if flow rate increases. A 2.6 µm core shell particle shows as same column efficiency as a totally porous sub-2 μm

The right figure shows that a diffusion width of a sample in a 2.6 µm core shell particle and a 2 µm totally porous particle. Both diffusion widths are almost same. The 2.6 um core shell particle is superficially porous, so that the diffusion width becomes narrower than particle size. Same diffusion means same efficiency.



Comparison of Performance by Plate/Pressure

	Plate	Back press. (MPa)	Plate/back press.
Sunniest C18 –HT 2.0 μm	9,900	16.7	593
Brand A C18 1.9 μm	7,660	16.3	470
Brand B C18 1.8 μm	10,100	19.6	515
Brand C C18 1.7 μm	11,140	32.0	348
SunShell C18 2.6 μm	9,600	9.7	990
Sunniest C18 –HT 2.0 µm Brand A C18 1.9 µm Brand B C18 1.8 µm Brand C C18 1.7 µm SunShell C18 2.6 µm			
	0 5,000 10,000	0 10 20 30	0 250 500 750 1000

Under a constant back pressure condition, SunShell C18 showed more than 2 times higher performance to compare with totally sub-2µm porous C18s.



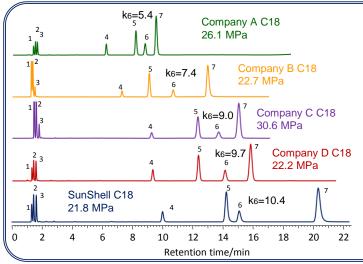
SunShell C18

Characteristics of SunShell C18

		Core shell	silica				C18		
	Particle size	Pore diameter	Specific surface area	I USP L# I End-capping I				Available pH range	
SunShell C18	2.6 μm	9 nm	150 m ² /g	7 %	C18	C18 L1 Sunniest End-capping 60 MPa or 8,570 psi		1.5 - 10	



Comparison of standard samples between core shell C18s



Company A C18, $150 \times 4.6 \text{ mm}$ (26.1 MPa) Company B C18, $150 \times 4.6 \text{ mm}$ (22.7 MPa) Company C C18, $150 \times 4.6 \text{ mm}$ (30.6 MPa) Company D C18, $150 \times 4.6 \text{ mm}$ (22.2 MPa) SunShell C18, $150 \times 4.6 \text{ mm}$ (21.8 MPa)

Mobile phase: CH₃OH/H₂O=75/25

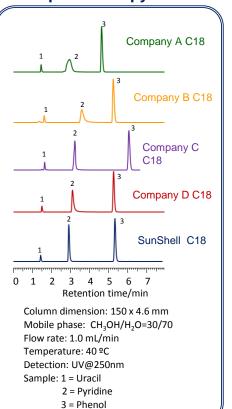
Flow rate: 1.0 mL/min, Temperature: 40 ºC

Sample: 1 = Uracil, 2 = Caffeine, 3 = Phenol, 4 = Butylbenzene 5 = o-Terphenyl, 6 = Amylbenzene, 7 = Triphenylene

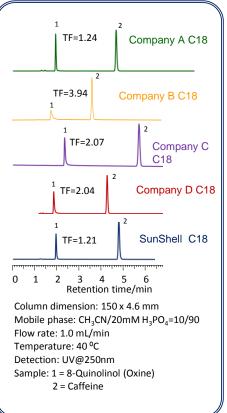
	Hydrogen bonding (Caffeine/Phenol)	Hydrophobicity (Amylbenzene/Butylbenzene)	Steric selectivity (Triphenylene/o-Terphenyl)
Company A C18	0.48	1.54	1.20
Company B C18	0.35	1.56	1.50
Company C C18	0.42	1.57	1.25
Company D C18	0.44	1.60	1.31
SunShell C18	0.39	1.60	1.46

Retention of standard samples and back pressure were compared for five kinds of core shell type C18s. Company A C18 showed only a half retention to compare with SunShell C18. Steric selectivity becomes large when ligand density on the surface is high. SunShell C18 has the largest steric selectivity so that it has the highest ligand density. This leads the longest retention time.

Comparison of pyridine

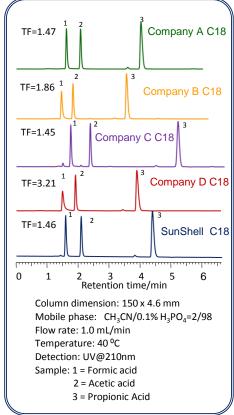


Comparison of Oxine



8-Quinolinol (Oxine) is a metal chelating compound. Metal impurities in the core shell particle leads the tailing for oxine peak.

Comparison of formic acid



Residual silanol groups make pyridine be tailing under methanol/water mobile phase condition. SunShell C18 shows a sharp peak for pyridine.

Formic acid is used as an indicator for a acidic inertness. SunShell and Company A and C C18 show a sharp peak.

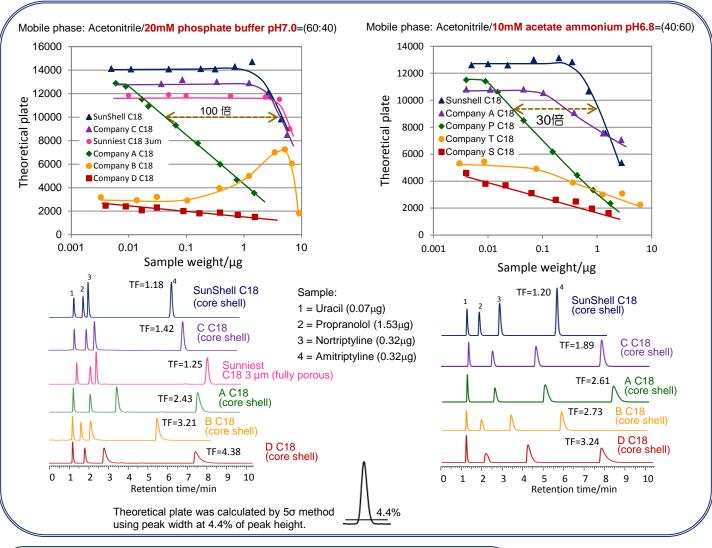


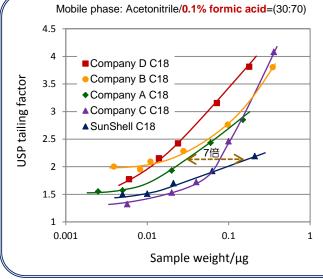
Loading capacity of amitriptyline as a basic compound

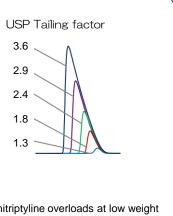
Amitriptyline overlords much more at acetonitrile/buffer mobile phase than methanol/buffer. Three kinds of core shell C18s were compared loading capacity of amitriptyline at three different mobile phases.

Common condition: Column dimension, 150 x 4.6 mm, flow rate; 1.0 mL/min, temperature; 40 °C









Amitriptyline overloads at low weight when acetonitrile/0.1% formic acid mobile phase. A peak is shifted forward under overloading.

Comparison column

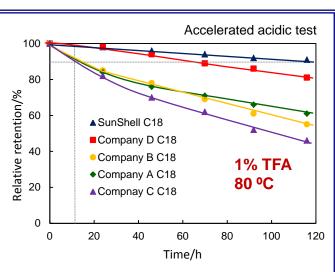
- 1. Kinetex C18, 2.6 μm
- 2. Accucore C18, 2.6 µm
- 3. PoroShell C18 EC, 2.7 µm
- 4. Ascentis Express C18, 2.7 μm
- 5. SunShell C18, 2.6 μm



All columns are core shell type. All columns sized 150 x 4.6 mm show 38,000 to 40,000 plates for a neutral compound. However regarding a basic compound like amitriptyline, SunShell C18 and company C C18 showed a good peak, while Company A, B and D C18 showed a poor peak. Company A C18 overloaded at more than 0.01 µg of amitriptyline while SunShell C18 overloaded at more than from 0.3 to 1 µg of amitriptyline. Surprisingly loading capacity of company A C18 was only one hundredth to compare with SunShell C18 under acetonitrile/20mM phosphate buffer pH7.0=(60:40) mobile phase. Company D C18 always showed poor peak of amitriptyline.



◆Evaluation of Stability



Durable test condition

Column size: 50 x 2.1 mm

Mobile phase: CH₃CN/1.0% TFA, pH1=10/90

Flow rate: 0.4 mL/min Temperature: 80 °C

Measurement condition

Column size: 50 x 2.1 mm Mobile phase: CH₃CN/H₂O=60/40

Flow rate: 0.4 mL/min Temperature: 40 °C Sample: 1 = Uracil (t_0) 2 = Butylbenzene

Stability under acidic pH condition was evaluated at 80 °C using acetonitrile/1% trifluoroacetic acid solution (10:90) as mobile phase. 100% aqueous mobile phase expels from the pore of packing materials by capillarity and packing materials doesn't deteriorate. 10% acetonitrile in a mobile phase allows an accurate evaluation.¹⁻³⁾

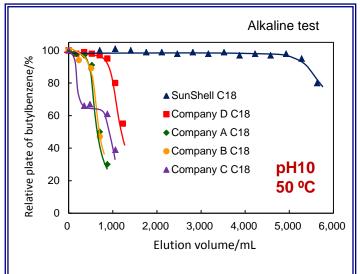
★Sunshell C18 has kept 90% retention for 100 hours under such a severe condition. SunShell C18 is 5 to 10 times more stable than the other core shell C18.

- 1) N. Nagae, T. Enami and S. Doshi, LC/GC North America October 2002.
- 2) T. Enami and N. Nagae, American Laboratory October 2004.
- 3) T. Enami and N. Nagae, BUNSEKI KAGAKU, 53 (2004) 1309.

Comparison column

- 1. Kinetex C18, 2.6 μm
- 2. Accucore C18, 2.6 μm
- 3. PoroShell C18 EC, 2.7 µm
- 4. Ascentis Express C18, 2.7 μm
- 5. SunShell C18, 2.6 μm





Durable test condition

Column Size: 50 x 2.1 mm

Mobile phase:

CH₃OH/20mM Sodium borate/10mM NaOH=30/21/49 (pH10)

Flow rate: 0.4 mL/min Temperature: 50 °C

Measurement condition

Column Size: 50 x 2.1 mm Mobile phase: CH₃CN/H₂O=60/40

Flow rate: 0.4 mL/min Temperature: 40 °C Sample: 1 = Butylbenzene

Stability under basic pH condition was evaluated at 50 °C using methanol/Sodium borate buffer pH 10 (30:70) as mobile phase. Sodium borate is used as a alkaline standard solution for pH meter, so that its buffer capacity is high.

Elevated temperature of 10 °C makes column life be one third. The other company shows stability test at ambient (room temperature). If room temperature is 25 °C, column life at room temperature (25 °C) is sixteen times longer than that at 50 °C.

- ★ SunShell C18 is enough stable even if it is used under pH 10 condition. Regarding stability under basic pH condition, there is little C18 column like SunShell C18 except for hybrid type C18. It is considered that our end-capping technique leads high stability.
- ★ SunShell C18 can be used at the pH range from 1.5 to 10.

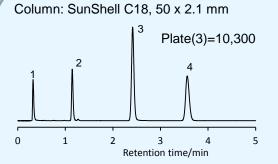




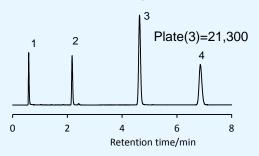
Efficiency of SunShell C18



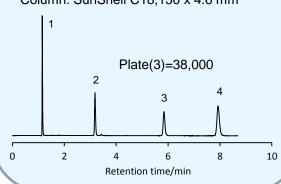
UHPLC



Column: SunShell C18, 100 x 2.1 mm



Column: SunShell C18,150 x 4.6 mm



Column: SunShell C18, 2.6 μm 50 x 2.1 mm

Mobile phase: CH₃CN/H₂O=60/40

Flow rate: 0.3 mL/min Pressure: 7 MPa Temperature: 23 °C

> Sample: 1 = Uracil 2 = Toluene 3 = Acenaphthene 4 = Butylbenzene

Column: SunShell C18, 2.6 µm 100 x 2.1 mm

Mobile phase: CH₃CN/H₂O=60/40

Flow rate: 0.3 mL/min Pressure: 12.5 MPa Temperature: 25 °C

Efficiency=253,000 plate/m

Column: SunShell C18, 2.6 μm $\,$ 150 x 4.6 mm $\,$ SunShell C18, 2.6 μm $\,$ 100 x 4.6 mm

Mobile phase: CH₃CN/H₂O=70/30

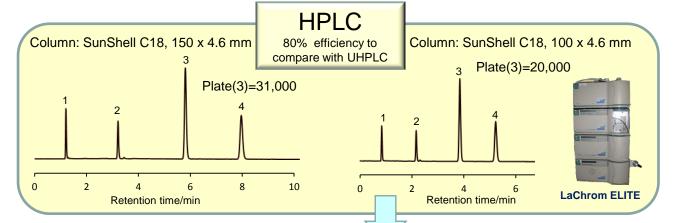
4 = Butylbenzene

Flow rate: 1.0 mL/min

Pressure: 14.5MPa(UHPLC),13.5 MPa(HPLC) for 150 mm

9.5MPa(HPLC) for 100 mm

Temperature: 25 °C
Sample: 1 = Uracil
2 = Toluene
3 = Acenaphthene



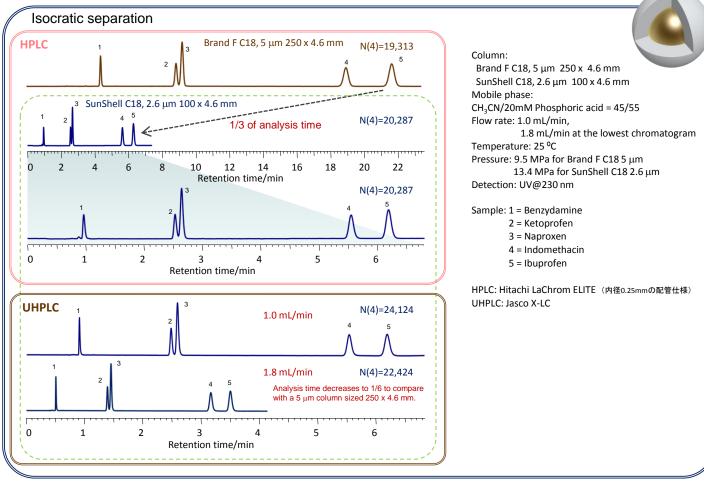


The same efficiency as 5 μm, 250 x 4.6 mm



Saving 60% for analytical time and consumption of solvent

Examples of transfer from a conventional 5 µm column to SunShell column



Comparison between normal and semi-micro HPLC

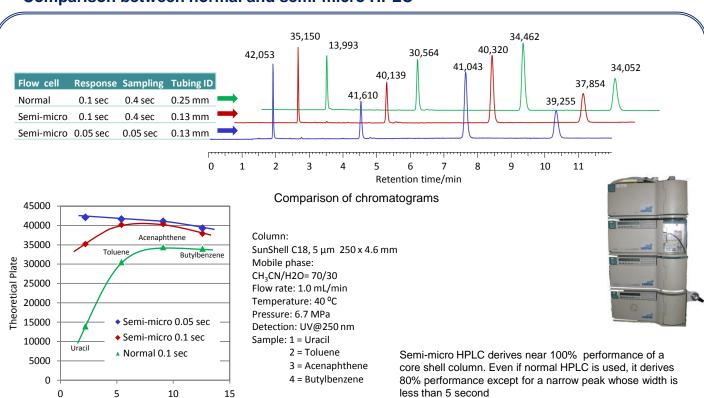
0

theoretical plate

5

Peak width/sec Relationship between Peak width and

10



HPLC: Hitachi LaChrom ELITE



SunShell C18-WP, RP-AQUA, C8, Phenyl, PFP

(Pentafluoropheny)

Characteristics of SunShell

		ore shell:	silica						
		Pore diameter	Specific surface area	Carbon content	Bonded phase	USP L#	End-capping	Maximum operating pressure	Available pH range
SunShell C18	2.6 μm	9nm	150 m ² /g	7%	C18	L1	Sunniest endcapping	60 MPa	1.5 - 10
SunShell C18-WP	2.6 μm	16 nm	90 m²/g	5%	C18	L1	Sunniest endcapping	60 MPa	1.5 - 10
SunShell RP-AQUA	2.6 μm	16 nm	90 m²/g	4%	C28	L62	Sunniest endcapping	60 MPa	2 - 8 ^{a)}
SunShell C8	2.6 μm	9nm	150 m ² /g	4.5%	C8	L7	Sunniest endcapping	60 MPa	1.5 - 9
SunShell Phenyl	2.6 μm	9nm	150 m ² /g	5%	Phenylhexyl	L11	Sunniest endcapping	60 MPa	1.5 - 9
SunShell PFP	2.6 μm	9nm	150 m ² /g	4.5%	Pentafluorophenyl	L43	TMS endcapping	60 MPa	2 - 8

♦ Comparison of standard samples

SunShell PFP

SunShell C8

12 14

Retention time/min

SunShell Phenyl

SunShell RP-AQUA

16 18

SunShell C18-WP

20 22

a) Under 100% aqueous condition

Column: SunShell C18, C18-WP, RP-AQUA, C8, Phenyl, PFP, 2.6 μm 150 x 4.6 mm

Mobile phase: CH₃OH/H₂O=75/25

Flow rate: 1.0 mL/min Temperature: 40 °C

Sample: 1 = Uracil

2 = Caffeine 3 = Phenol

4 = Butylbenzene

5 = o-Terphenyl

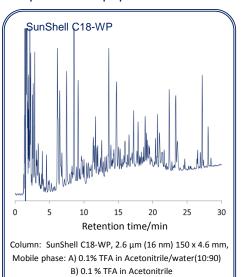
6 = Amylk 7 = Triphe		\rightarrow	
	Hydrogen bonding (Caffeine/Phenol)	Hydrophobicity (Amylbenzene/Butylbenzene)	Steric selectivity (Triphenylene/o-Terphenyl)
PFP	1.00	1.31	2.38
Phenyl	1.00	1.48	1.01
C8	0.32	1.46	1.08
RP-AQUA	0.52	1.52	1.30
C18-WP	0.40	1.55	1.35
SunShell C18	0.39	1.60	1.46

Separation of peptides

SunShell C18

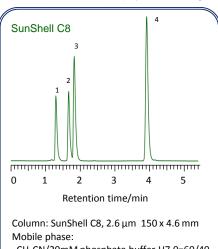
6

8 10



Gradient program: %B 5% – 50% in 50 min
Flow rate: 1.0 mL/min , Temperature: 25 °C,
Detection: UV@210 nm,
Sample: Tryptic digest of BSA

Separation of amitriptyline using C8



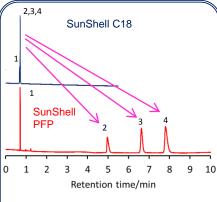
CH₃CN/20mM phosphate buffer H7.0=60/40

Flow rate: 1.0 mL/min Temperature: 40 °C Detection: UV@250nm

Sample: 1 = Uracil, 2 = Propranolol,

3 = Nortriptyline, 4 = Amitriptyline

Separation of basic compounds



Column: SunShell C18, 2.6 μm 150 x 4.6 mm SunShell PFP, 2.6 μm 150 x 4.6 mm

Mobile phase:

CH₃CN/10mM phosphate buffer pH7.0=80/20

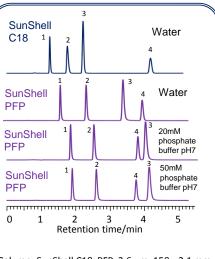
Flow rate: 1.8 mL/min Temperature: 25 °C

Sample: 1 = Uracil, 2 = Propranolol,

3 = Nortriptyline, 4 = Amitriptyline



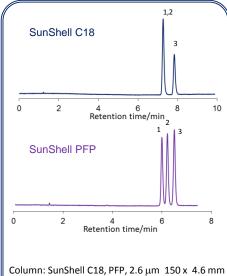
Separation of xanthines



Column: SunShell C18, PFP, 2.6 µm 150 x 2.1 mm Mobile phase: CH₃OH/water or buffer=30/70

Flow rate: 0.3 mL/min Temperature: 25 °C Detection: UV@250nm Sample: 1 = Theobromine 2 = Theophyline 3 = Caffeine 4 = Phenol

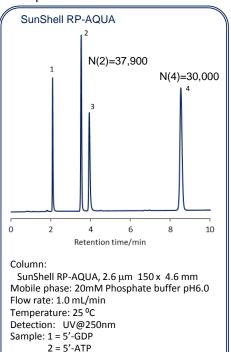
Separation of cresol isomers



Mobile phase: CH₃OH/H₂O=40/60

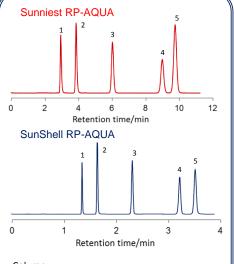
Flow rate: 1.0 mL/min Temperature: 25 °C Detection: UV@250nm Sample: 1 = p-Cresol 2 = m-Cresol 3 = o-Cresol

Separation of nucleotides



3 = 5' - ADP4 = 5' - AMP

Separation of nucleic acid bases



Sunniest RP-AQUA, 5 μm 150 x 4.6 mm SunShell RP-AQUA, 2.6 µm 150 x 4.6 mm Mobile phase:

10mM Phosphate buffer pH7.0 Flow rate: 1.0 mL/min for Sunniest

1.5 ml/min for SunShell

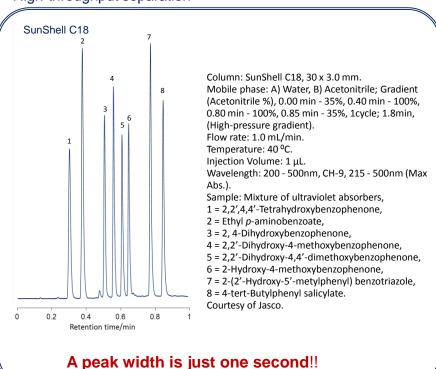
Temperature: 24 °C

Sample: 1 = Cytosine, 2 = Uracil, 3 = Thymidine,

4 = Uridine, 5 = Thymine

	Plate(5)	Resolution (4,5)
Sunniest	14,000	1.98
SunShell	30,000	3.79

High-throughput separation







SunShell HFC18-16, HFC18-30

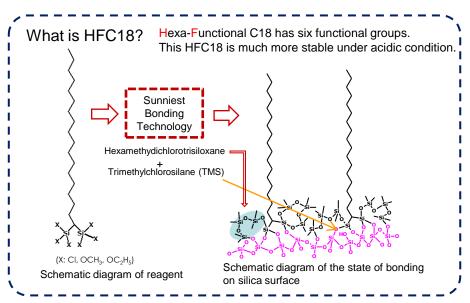


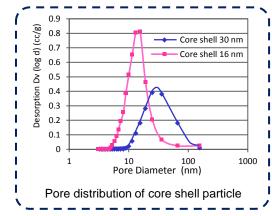
Characteristics of SunShell HFC18

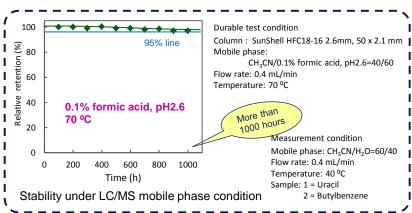
For separation of peptides and proteins

		Core shel	l silica		C18 (USP L# 1)						
	Particl e size	Pore diameter	Specific surface area	Carbon content	Ligand density	End-capping	Maximum operating pressure	Available pH range			
SunShell C18-WP	2.6 μm	16 nm	90 m²/g	5 %	2.5 μmol/m2	Sunniest endcapping	60 MPa or 8,570 psi	1.5 - 10			
SunShell HFC18-16	2.6 μm	16 nm	90 m²/g	2.5%	1.2 μmol/m2	Sunniest endcapping	60 MPa or 8,570 psi	1.5 – 9			
SunShell HFC18-30	2.6 μm	30 nm	40 m²/g	1.3%	1.2 μmol/m2	Sunniest endcapping	60 MPa ^a or 8,570 psi ^a	1.5 - 9			

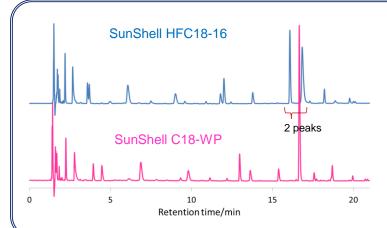
a: 50MPa, 7141psi for 4.6 mm i.d. column







Separation of peptides



Column: SunShell HFC18-16, 2.6 µm (16 nm) 150 x 4.6 mm, SunShell C18-WP, 2.6 µm (16 nm) 150 x 4.6 mm Mobile phase: A) 0.1% TFA in Acetonitrile/water(10:90) B) 0.1 % TFA in Acetonitrile

Gradient program: Time 0 min 5 mi

%B 5% 5% 5

Flow rate: 1.0 mL/min Temperature: 25 °C Detection: UV@210 nm

Sample: Tryptic digest of cytochrome C



SunShell 2-EP

For Supercritical fluid Chromatography

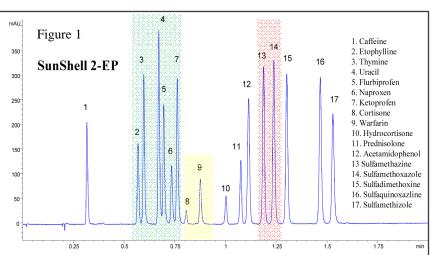
2.6 μ m core shell column shows only one third of back pressure to compare with 1.7 μ m fully porous column although both show almost same efficiency. By such low back pressure, a difference of density of supercritical fluid between an inlet and an outlet of the column is reduced. Consequently, . 2.6 μ m core shell column performs a superior separation for SFC.

Characteristics of SunShell 2-EP

Ì			Core shell sili	ca					
		Particle size	Pore diameter	Specific surface area	Carbon content	Bonded phase	End- capping	Maximum operating pressure	Available pH range
	SunShell 2-EP	2.6 μm	9 nm	150 m²/g	2.5%	2-Ethylpyridine	no	60 MPa or 8,570 psi	2 – 7.5

Comparison between SunShell 2-EP and 1.7 µm fully porous 2-EP





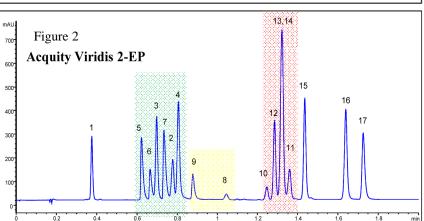
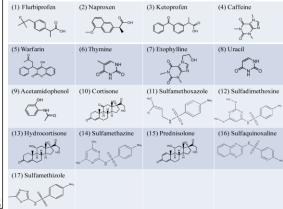


Figure 1: Chromatogram of the separation for he 17-component mix using the Sun Shell 2-EP 150 x 3.0 mm column. A methanol gradient of < 2 minutes was used on the Agilent 1260 Infinity SFC system. SFC conditions: flow rate: 4.0mL/min; outlet pressure 160 bar; column temperature 55 $^{\circ}$ C. Gradient program: 5.0-7.5% in 0.20 min, then 7.5-20% in 1.3 min and held at 20% for 0.2 min.

Figure 2: Chromatogram of the separation for the 17-component mix using Acquity UPC² Viridis 2-EP 100 x 3.0 mm column. 16 of the 17 components were resolved. A methanol gradient of < 2 minutes was used on the Agilent 1260 Infinity SFC system. SFC conditions: flow rate 3.5 mL/min; outlet pressure 160 bar; and column temperature 70° C. Gradient program: 5.0-12.5% in 1.0 min, 12.5% for 0.25 min, then 12.5-20% in 0.75 min.



Courtesy of Pfizer Inc.



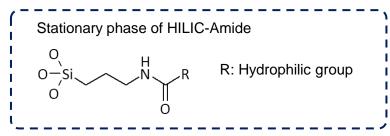


SunShell HILIC-Amide

For Hydrophilic Interaction Chromatography

Characteristics of SunShell HILIC-Amide

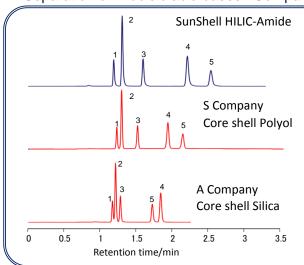
		Core shell sil	ica			Amide (l	JSP L# 68)	
	Particle size	Pore diameter	Specific surface area	I End-capping I		Available pH range		
SunShell HILIC-Amide	2.6 μm	9 nm	150 m²/g	3%	Amide	no	60 MPa or 8,570 psi	2 - 8





Stationary phase of SunShell HILIC-Amide consists of AMIDE and HYDROPHILIC GROUP, so that this stationary phase is more polar than an individual group. High speed separation is leaded by core shell structure that derives high efficiency and fast equilibration.

Separation of Nucleic acid bases: Comparison of the other core shell hilic columns



Column:

SunShell HILIC-Amide, 2.6 μ m 100 x 4.6 mm, Coreshell polyol, 2.7 μ m 100 x 4.6 mm, Core shell Silica, 2.7 μ m 100 x 4.6 mm Mobile phase:

Mobile phase:

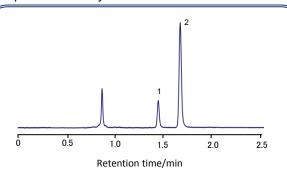
Acetonitrile/20 mM ammonium acetate(pH4.7) = 8/2

Flow rate: 1.0 mL/min Temperature: 40 °C Detection: UV@250 nm

Sample: 1 = Thymine, 2 = Uracil, 3 = Uridine, 4 = Cytosine, 5 = Cytidine

Regarding retention of cytidine, SunShell HILIC-Amide showed 30% higher retention factor than S core shell polyol.

Separation of Cyanuric acid and Melamine



Column: SunShell HILIC-Amide, 2.6 μm 100 x 4.6 mm

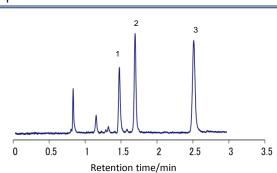
Mobile phase:

Acetonitrile/5 mM phosphate Buffer (pH6.9) =75/25

Flow rate: 1.0 mL/min Temperature: 40 °C Detection: UV@220 nm,

Sample: 1 = Cyanuric acid, 2 = Melamine

Separation of water- soluble vitamins



Column: SunShell HILIC-Amide, 2.6 µm 100 x 4.6 mm

Mobile phase:

Acetonitrile/25 mM phosphate buffer (pH2.5) =8/2

Flow rate: 1.0 mL/min Temperature: 40 °C Detection: UV@250 nm,

Sample: 1 = Nicotinic acid, 2 = Ascorbic acid, 3 = Pyridoxine



SunShell RP Guard Filter

< Cartridge Type, Bonded with C18 and End-Capped with TMS>

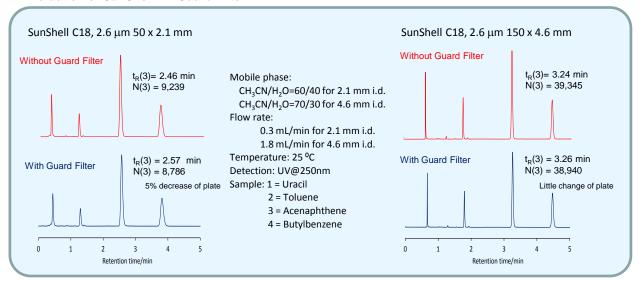
Available as a guard column for reversed phase





- √ The filter is made of porous glass sized 4 mm i.d. and 4 mm thickness.
- ✓ Pore diameter is 2 μm.
- ✓ Low dead volume structure
- ✓ Back pressure on glass filter is ca. 0.1 MPa at 1.0 mL/min of flow rate.
- ✓ Upper pressure limit is more than 60 MPa
- ✓ Available for 2.1 mm i.d to 4.6 mm i.d. column

Evaluation of SunShell RP Guard Filter



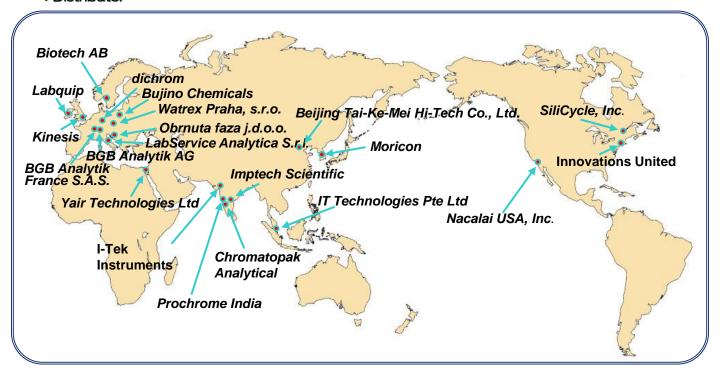
Price of SunShell RP Guard Filter

Name	quantity	Part number	Photo
SunShell RP Guard Filter For exchange	5 pieces	CBGAAC	
SunShell RP Guard Filter Holder	1 piece	CBGAAH	





*Distributor





Ordering information of SunShell 2.6 μm

	Inner diameter (mm)	1.0	2.1	3.0	4.6	USP category	
	Length (mm)	Catalog number	Catalog number	Catalog number	Catalog number		
	30		CB6931	CB6331	CB6431		
	50	CB6141	CB6941	CB6341	CB6441		
SunShell C18	75		CB6951	CB6351	CB6451	L1	
	100	CB6161	CB6961	CB6361	CB6461		
	150	CB6171	CB6971	CB6371	CB6471		
	30		CC6931	CC6331	CC6431		
	50		CC6941	CC6341	CC6441		
SunShell C8	75		CC6951	CC6351	CC6451	L7	
	100		CC6961	CC6361	CC6461		
	150		CC6971	CC6371	CC6471		
	30		CF6931	CF6331	CF6431		
	50		CF6941	CF6341	CF6441		
SunShell PFP	75		CF6951	CF6351	CF6451	L43	
	100		CF6961	CF6361	CF6461		
	150		CF6971	CF6371	CF6471		
	30		CW6931	CW6331	CW6431		
	50		CW6941	CW6341	CW6441		
SunShell C18-WP	75		CW6951	CW6351	CW6451	L1	
	100		CW6961	CW6361	CW6461		
	150		CW6971	CW6371	CW6471		
	30		CR6931	CR6331	CR6431		
	50	CR6141	CR6941	CR6341	CR6441		
SunShell RP-AQUA	75		CR6951	CR6351	CR6451	L62	
	100	CR6161	CR6961	CR6361	CR6461		
	150	CR6171	CR6971	CR6371	CR6471		
	30		CP6931	CP6331	CP6431		
	50		CP6941	CP6341	CP6441		
SunShell Phenyl	75		CP6951	CP6351	CP6451	L11	
,	100		CP6961	CP6361	CP6461		
	150		CP6971	CP6371	CP6471		
	30		CH6931	CH6331	CH6431		
	50		CH6941	CH6341	CH6441		
SunShell HILIC-Amide	75		CH6951	CH6351	CH6451	L68	
	100		CH6961	CH6361	CH6461		
	150		CH6971	CH6371	CH6471		
	30		CE6931	CE6331	CE6431		
	50		CE6941	CE6341	CE6441		
SunShell 2-EP	75		CE6951	CE6351	CE6451		
	100		CE6961	CE6361	CE6461		
	150		CE6971	CE6371	CE6471		
	50		CG6941	CG6341	CG6441		
SunShell HFC18-16	100		CG6961	CG6361	CG6461	L1	
	150		CG6971	CG6371	CG6471		
	50		C46941	C46341	C46441		
SunShell HFC18-30	100		C46961	C46361	C46461	L1	
	150		C46971	C46371	C46471		



Manufacturer

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